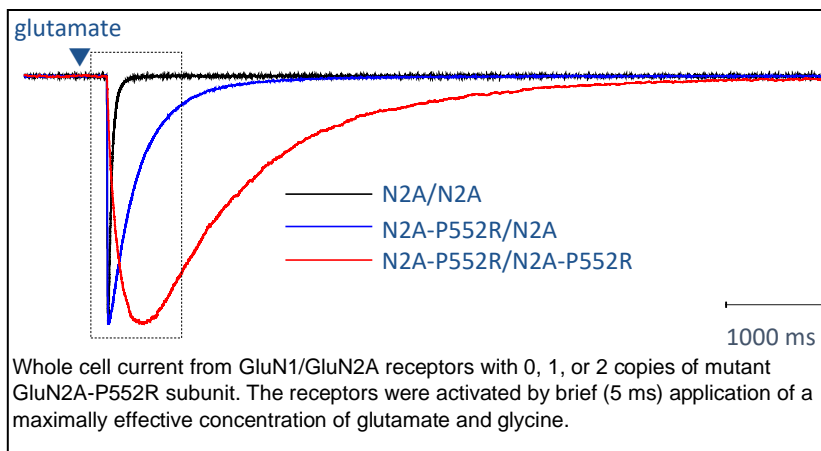
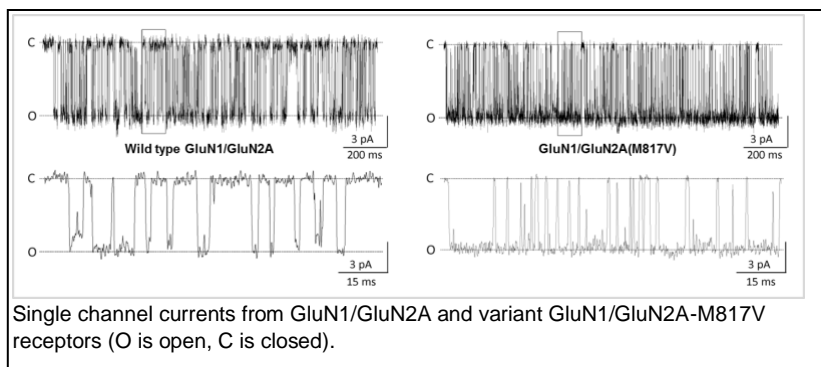


Functional Effects of Glutamate Receptor Mutations in Human Diseases

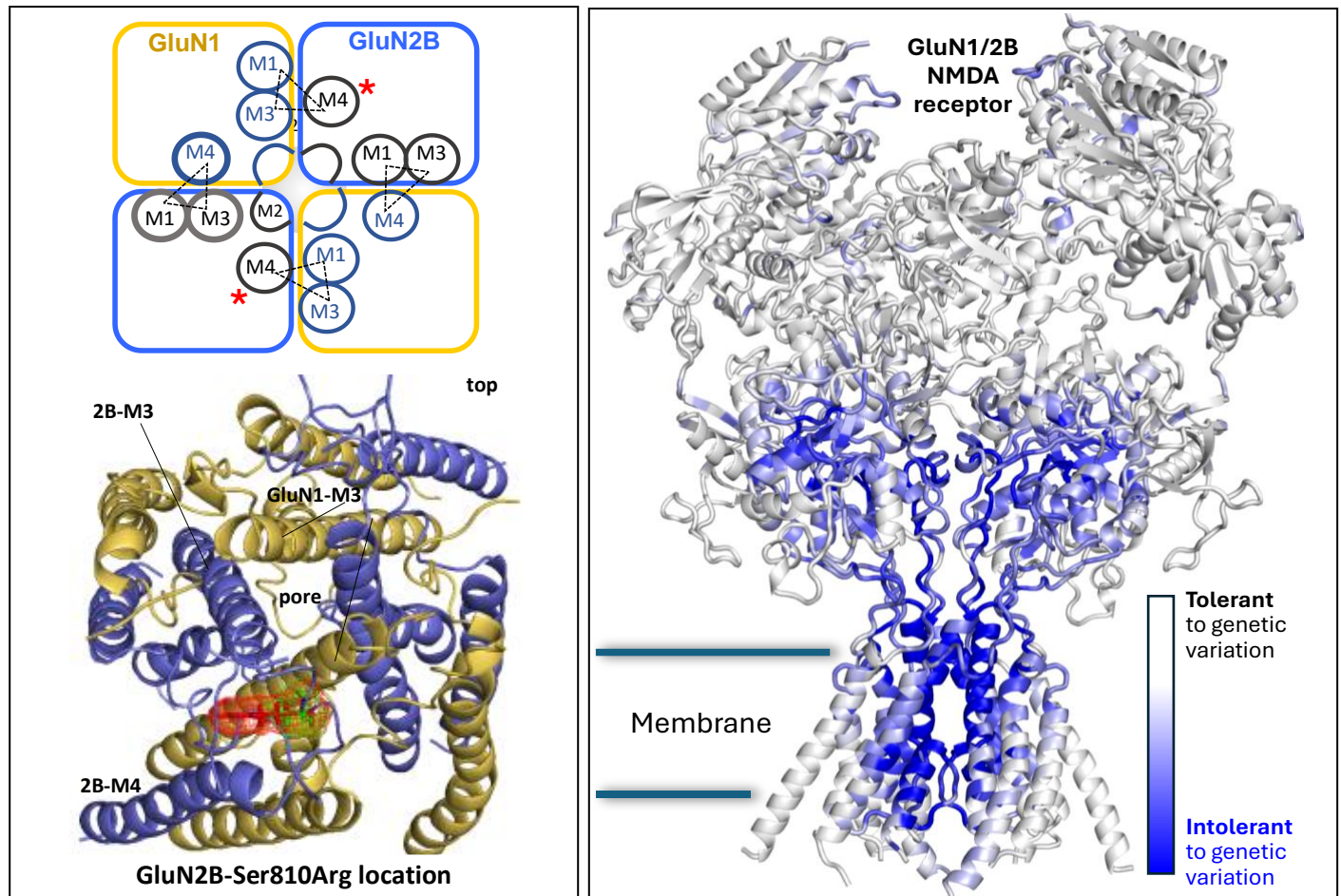
Many early life disorders, including epilepsy and developmental delay, can involve genetic errors, most often *de novo* mutations that produce missense variants. Genome or exome DNA sequencing can provide pediatric patients who experience these disorders with definitive diagnoses, reducing unnecessary testing, bringing clarity to the family, and focusing basic and clinical research resources. These diagnoses have yielded clinical insight showing that the majority of epilepsies that start prior to one year of life (e.g. infantile epileptic encephalopathies) occur due to *de novo* genetic errors. However, a large gap still remains between the genetic information describing missense variants in patients and our understanding of how these variants affect the function of the proteins encoded by the affected gene. This lack of functional understanding prevents the translation of genetic information into a better mechanistic understanding and treatment of disease. We are working to directly solve this problem.

Multiple lines of evidence support the idea that genetic variants in genes encoding ion channels can give rise to neurological disease. It is now clear that *de novo* variants in many of the 18 genes encoding glutamate receptor subunits are a major contributor to channelopathies--diseases arising from altered expression or function of an ion channel. We work in partnership with the Center for Functional Evaluation of Rare Variants at Emory and the Simon's Foundation to elucidate the functional consequences of all *de novo* NMDA receptor variants, which exceed 1000 in number. Many of these variants profoundly alter receptor function. We are working to establish the relationship between allelic frequency and functional effects to establish diagnostic criteria for patients with rare NMDA receptor variants. In addition, we are analyzing our functional data for hundreds of variants in the context of clinical data to explore the mechanisms underlying these conditions. All functional data we generate is placed on a public database (the GRIN Portal) hosted by the Broad Institute.

In addition, the functional information we obtain is being used by clinicians to stratify patients for ongoing and future clinical trials, which helps to diminish patient heterogeneity and increase the power of these smaller trials for rare diseases. This is essential given the low number of available patients to enroll. This information also can serve as a training set for machine learning algorithms optimized to predict the functional consequences of future variants, and further expand information that can be ascertained from future sequencing.



Lastly, functional data has revealed in exquisite detail the structural basis of ion channel function since many variants have been identified in regions previously not known to be involved in gating. This work has further validated the predictive power of the analysis of intolerant regions within the gene, which show a reduced number of variants in the healthy population. These intolerant regions are identified through algorithms that we helped to develop. Our functional data show that the regions that are predicted to be intolerant harbor a large number of variants.



GluN2A pre-M1 and M3 plus GluN1 pre-M4 may interact to control gating

>300 *de novo* mutations in NMDARs, primarily concentrated in gating domains

M3/SYTANLAAF

GluN1-G638V
GluN1-M641I
GluN1-A645S
GluN1-Y647C
GluN1-Y647S
GluN1-N650K
GluN1-N650I
GluN1-A652T
GluN1-A653G
GluN1-L655Q
GluN2A-L642M
GluN2A-A643D
GluN2A-S644G
GluN2A-T646A
GluN2A-N648S

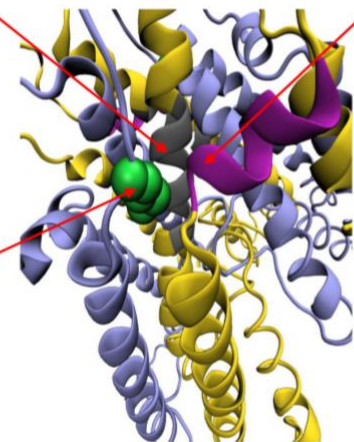
GluN2A-L649V
GluN2A-F652V
GluN2B-A636P
GluN2B-A636V
GluN2B-A639V
GluN2B-I641T
GluN2B-N649T
GluN2B-A652G
GluN2B-M653I
GluN2B-M653V
GluN2B-I655F
GluN2D-A663P
GluN2D-V667I
GluN2D-A675T
GluN2D-A678D
GluN2D-M681I

Pre-M1

GluN1-S547del
GluN1-S549G
GluN1-S549R
GluN1-L551P
GluN1-D552E
GluN1-Q556
GluN1-P557R
GluN1-S560dup
GluN2A-S545L
GluN2A-S547del
GluN2A-A548T
GluN2A-P552R
GluN2B-S541R
GluN2B-S555I
GluN2B-G543R
GluN2B-P553L
GluN2B-P553T

Pre-M4

GluN1-A806T
GluN1-G815R
GluN1-G815V
GluN1-G827R
GluN1-F817L
GluN2A-L812M
GluN2A-I814T
GluN2A-M817V
GluN2A-M817T
GluN2A-M818L
GluN2B-E807R
GluN2B-S810N
GluN2B-S810R
GluN2B-M818T
GluN2B-A819T
GluN2B-G820A
GluN2B-G820E
GluN2B-G820V
GluN2B-M824R
GluN2B-L825V
GluN2B-G826E



23% of *de novo* mutations occur in <2.5% of protein

CFEY CENTER FOR FUNCTIONAL EVALUATION OF RARE VARIANTS

25% of missense variants reside in the pre-M1 two-turn helix (magenta), pre-M4 linker (green), and M3-SYTANLAAF motif (charcoal). GluN1 is yellow and